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1 Cellulose gels produced in room temperature ionic liquids
2 by ionizing radiation

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11
12 **Abstract**

13 Cellulose-based gels were produced in room temperature ionic liquids (RTILs)
14 by ionizing radiation. Cellulose was dissolved at the initial concentration of 20 wt.% in
15 1-ethyl-3-methylimidazolium (EMI)-acetate or *N,N*-diethyl-*N*-methyl-
16 *N*-(2-methoxyethyl)ammonium (DEMA)-formate with a water content of 18 wt.%, and
17 irradiated with γ -rays under aerated condition to produce new cellulose gels. The gel
18 fractions of the cellulose gels obtained in EMI-acetate and DEMA-formate at a dose of
19 10 kGy were 13 % and 19 %, respectively. The formation of gel fractions was found to
20 depend on the initial concentration of cellulose, water content, and irradiation

21 temperature. The obtained gel readily absorbed water, methanol, ethanol,
22 dichloromethane, *N, N*-dimethylacetamide, and RTILs.

23

24 **Highlight**

- 25 • Cellulose gels were produced in room temperature ionic liquids (RTILs).
- 26 • Water plays a crucial role in the cross-linking reaction.
- 27 • Cellulose gels swollen with RTILs show good electronic conductivity (3.0
28 $\text{mS}\cdot\text{cm}^{-1}$).

29

30 **Keywords**

31 cellulose, gel, room temperature ionic liquid, ionizing radiation.

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39 **1. Introduction**

40 In recent years, natural polymers have attracted significant attention as alternative
41 green materials because of the exhaustion of petroleum and non-renewable resources,
42 and the increase in environmental pollution. Polysaccharides such as cellulose,
43 chitin/chitosan, starch, and alginic acid are natural polymers. Large quantities of
44 polysaccharide-based biomass are ubiquitous on Earth. They possess unique structures,
45 distinctive properties, and offer the advantages of being non-toxic and biodegradable.
46 They are often modified to produce functional materials, such as viscose rayon,
47 personal care products, and biodegradable plastics. Ionizing radiation is widely used as
48 a convenient tool for the modification of polysaccharides via decomposition, grafting,
49 and cross-linking (Seko et al., 2005). Besides, chemically modified polysaccharides
50 such as carboxymethyl cellulose, hydroxypropylmethyl cellulose, carboxymethyl
51 chitin/chitosan, and carboxymethyl starch, all of which dissolve in water at high
52 concentrations, have also been irradiated with ionizing radiation to obtain derivative
53 gels that are used as materials for making water absorbents and wound dressings (Fei
54 et al., 2000; Wach et al., 2001; Zhao et al., 2003; Zhao et al., 2008; Nagasawa et al.,
55 2004; Pekel et al., 2004; Wang et al., 2007). Until now, radiation cross-linked materials
56 have only been produced either from polysaccharide derivatives or using cross-linking

57 reagents, as certain natural polysaccharides have poor solubility in water and organic
58 solvents as well as low chemical reactivity (Kumar, 2000). These limitations could
59 possibly be circumvented by the use of organic solutions that have high
60 proton-accepting ability, such as chloroform in dimethylsulfoxide,
61 *N*-methylmorpholine-*N*-oxide, lithium chloride (LiCl) in *N,N*-dimethylacetamide,
62 calcium chloride in methanol, and room temperature ionic liquids (RTILs), which can
63 cleave the hydrogen bonds of the polysaccharides to form solutions in the
64 concentration range of 0.1–20 wt.% (Swatloski et al., 2002; Heinze et al., 2005;
65 Fukaya et al., 2008).

66 Among these organic solutions, RTILs have attracted much attention in the
67 diverse fields of chemistry, biology, engineering, and environmental science because of
68 their unique functionalities such as specific solubility, chemical and thermal stabilities,
69 unusual dynamical properties linked to high viscosity, and wide potential windows
70 (Rogers, 2003). So far, several studies have been reported in the fields of photo- and
71 radiation chemistry to identify more effective reactions between the charged reactive
72 species in RTILs (Wishart et al., 2003; Wishart et al., 2005; Kondoh et al., 2009; Katoh
73 et al., 2009; Kimura et al., 2010). In addition, extensive fundamental and applied
74 research have been conducted on RTILs used in various fields such as

75 radiation-induced polymerization (Qi et al., 2008) and radiation-induced grafting
76 polymerization (Hao et al., 2009). However, to the best of our knowledge, there are no
77 studies reported so far on the use of RTILs in radiation cross-linking reactions.

78 Therefore, in this study, we focus on the application of RTILs as “specific
79 solvents for the dissolution of polysaccharides,” and their use in establishing a “proper
80 reaction field for radiation cross-linking.” We propose a new method for the radiation
81 modification of polysaccharides using RTILs to produce new functional and
82 sustainable materials. Cellulose was selected as the polysaccharide of choice as it is the
83 most prevalent plant-derived polysaccharide, which comprises the main component of
84 plant plasma membranes and fibrous raw materials such as paper, wood material, and
85 cotton. Recently, it has been reported that the degree of polymerization of 5 wt.%
86 cellulose dissolved in 1-butyl-3-methylimidazolium (BMI) –chloride tends to reduce as
87 a function of dose (Hao et al. 2012). Meanwhile, the scission reaction is preferred over
88 the cross-linking reaction when the concentration of polysaccharide in the organic
89 solvent is less than approximately 10 wt.% (Tabata et al., 1991; Nagasawa et al., 2004).
90 We assume that cross-linked materials could be produced from cellulose via ionizing
91 radiation in RTILs, which are capable of dissolving more than 10 wt.% of cellulose.

92 We designed and synthesized some RTILs as the reaction medium, in which
93 cellulose could be dissolved to concentrations in excess of 10 wt.%, and further
94 investigated the radiation-induced cross-linking of cellulose in each RTIL. The
95 outcome of this study is expected to provide insights on new ways of functionalizing
96 materials, including cellulose and other natural polymers.

97

98 **2. Experimental**

99 *2.1 Sample preparation*

100 Cellulose powder (Microcrystalline grade, Merck) was used as received and its
101 viscosity-average molecular weight was estimated by the following
102 Mark–Kuhn–Houwink–Sakurada equation:

$$103 \quad [\eta] = K M_v^\alpha \quad (1)$$

104 where $[\eta]$ and M_v are the viscosity and viscosity-average molecular weight of the
105 sample solution, respectively, and K and α are constants reported as 1.278×10^{-4}
106 $\text{cm}^3 \cdot \text{g}^{-1}$ and 1.19, respectively (McCormick, 1985). In the typical process,
107 pre-determined quantity of cellulose was dissolved in the solution of 9 wt.% LiCl
108 (Wako, 98 %) in *N, N*-dimethylacetamide (DMAc, Wako, 97 %). The viscosity of the
109 cellulose solutions in the concentration range 1 - 4 $\text{g} \cdot \text{L}^{-1}$ was measured using Ostwald

110 and oscillatory viscometers (CBC Co., Ltd., Viscomate VM-10A). The weight-average
111 molecular weight of cellulose was analyzed using a high-performance liquid
112 chromatograph (HPLC, Shimadzu, SCL-10A) with gel permeation chromatography
113 (GPC) columns (TSL-GEL ALPHA and 2500, Tosoh) and a refractive index detector
114 (RI 704, GL Science). For HPLC measurement, the flow rate of the 30 mmol·L⁻¹
115 LiCl/DMAc eluent was maintained as 0.3 mL·min⁻¹, and the column temperature was
116 set at 313 K. Subsequently, 100 μL each of the prepared cellulose solutions was
117 injected to the HPLC. The standard for the GPC analysis was prepared by dissolving
118 1-4 g·L⁻¹ of polymethylmethacrylate (PMMA, Shodex, molecular mass in the range
119 1.8×10^3 – 2.0×10^5 g·mol⁻¹) in a solution of 0.2 wt.% LiCl/DMAc.

120 Cellulose was dissolved in each RTIL using a hybrid mixer (KEYENCE,
121 HM-500) to achieve initial concentrations in the range 1–30 wt.% and to obtain various
122 viscous and paste-like solutions. The RTILs used in this study, namely,
123 BMI-bis(trifluoromethylsulfonyl)amide (TFSA), BMI-thiocyanate, BMI-chloride,
124 diallylimidazolium (DAI)-chloride, *N,N*-diethyl-*N*-methyl-*N*-(2-methoxyethyl)
125 ammonium (DEMA)-chloride, and 1-ethyl-3-methylimidazolium (EMI)-bromide, were
126 purchased from Kanto Chemical Co., Inc., Japan. EMI-acetate, at 90 % concentration,
127 was also purchased from Aldrich Chemical Co., Inc. The other RTILs used in this study,

128 namely, BMI-acetate, DAI-formate, DAI-acetate, DEMA-formate, and DEMA-acetate
129 were synthesized by the methodology reported in the literature (Fukaya et al., 2008). In
130 the typical process, RTILs having halides, namely, BMI-chloride, DAI-chloride, and
131 DEMA-bromide, were used as precursors, which were converted into their respective
132 hydroxides using a column filled with an anion-exchange resin (Aldrich, Amberlite
133 IRA 400-OH). The obtained aqueous solutions of RTILs with a pH of 12 were
134 neutralized with either formic acid (Wako, 97 %) or acetic acid (Wako, 1 mol·L⁻¹
135 aqueous solution) to prepare RTILs functionalized with formate or acetate, respectively.
136 These RTILs were then washed with hexane (Wako, 96 %) and pure water (TOC: 4
137 ppb, electrical resistance: 18.2 MΩ·cm⁻¹) supplied from a Millipore Milli-Q system,
138 and vacuum dried overnight at 353 K. All the sample solutions were prepared in a
139 glove bag (Glass-Col) or glove box (Sanplatec) under dry or wet air saturated
140 conditions. The water content in the RTIL solutions was controlled by changing the
141 humidity of the surrounding air. This humidity was controlled by using a 0.1–2
142 mol·L⁻¹ aqueous solution of calcium chloride (Wako, 99 %) in both the bag and box
143 (Stokes, 1949). The water content in the cellulose solutions was measured with a
144 thermo-balance (Shimadzu AUW120D) and a Karl Fischer titration system (Kyoto
145 Electronics Manufacturing Co., Ltd., MKC-501).

146 2.2 ⁶⁰Co γ -ray irradiation

147 The cellulose solutions were irradiated using a ⁶⁰Co γ -ray source facility at the
148 Japan Atomic Energy Agency, Takasaki, in the dose range of 5–100 kGy (Gy = J·kg⁻¹)
149 with the dose rates ranging from 5–10 kGy·h⁻¹. The temperature of the sample
150 solutions was controlled using a thermostatic chamber (DTU-1C, TAITEC) during the
151 irradiation. Dosimetric experiments were carried out with an ethanol (Wako, > 99.7 %,
152 HPLC grade)–chlorobenzene (Aldrich, 99.9 %, HPLC grade) dosimeter and an
153 alanine-based dosimeter (Hitachi Cable, Ltd., AMINOGRAY, alanine 70 %,
154 polystyrene 30 %, volume: $\phi 3 \times 30$ mm) in a plastic container ($\phi 12 \times 50$ mm, 4 mm in
155 thickness) (Kojima et al., 1993).

156

157 2.3 Measurements

158 The irradiated sample solutions were subsequently washed with a solution of 5
159 wt.% LiCl/DMAc to remove any residues of unreacted cellulose, RTILs, and
160 degradation products. The residual matter was rinsed with water on a stainless steel
161 mesh (SUS316, 200 mesh) and then dried overnight at 353 K. The dried sample was
162 weighed in a thermo–balance. Rheological measurement of the sample solutions after
163 γ -ray irradiation was performed on a dynamic viscoelasticity measurement system (TA

164 instruments, ARES RDA-3) at 353 K, in the frequency range of 0.5–100 rad·s⁻¹.
165 Besides, the residual solutions containing unreacted cellulose, RTILs, and degradation
166 products were analyzed in the HPLC system with the GPC column and the refractive
167 index detector. Electrical conductivity of the samples was evaluated using a chemical
168 impedance analyzer (HIOKI, 3522-50) at the frequency of 100 kHz and temperature of
169 298 K.

170

171

172 **3. Results and Discussion**

173 *3.1 Radiation modification of cellulose in RTILs*

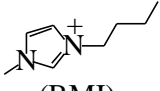
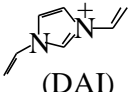
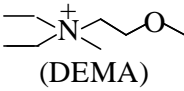
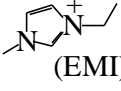
174 The weight-average molecular weight of cellulose before irradiation was 1.3×10^5
175 g·mol⁻¹, as determined using the Mark–Kuhn–Howink–Sakurada equation and GPC
176 analysis. The paste-like solutions of cellulose were prepared in each RTIL with initial
177 cellulose concentration in the range 10–30 wt.% at 298 K. As listed in Table 1, the
178 concentration of cellulose dissolved in carboxylate–based RTILs was found to be
179 higher than those dissolved in other RTILs.

180

181

182

183 Table 1 Solubility and gel fraction of cellulose mixed in room temperature ionic liquids
 184 under dehydrated condition at 298 K.

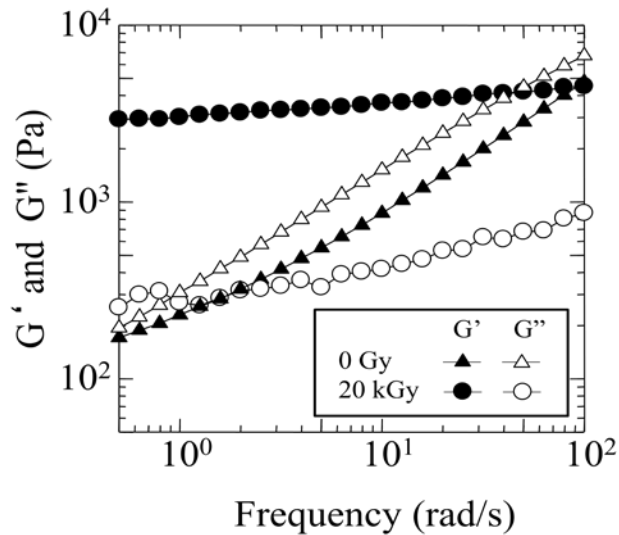
Cation	Anion	Concentration of cellulose in paste-like solution (wt.%)
 (BMI)	bis(trifluoromethylsulfonyl)amide	Insoluble
	thiocyanate	Insoluble
	chloride	10
	formate	20
 (DAI)	formate	20
	acetate	20
 (DEMA)	chloride	10
	formate	20
	acetate	20
 (EMI)	acetate	20–30

185 The paste-like solution obtained by mixing 20 wt.% cellulose in EMI-acetate under
 186 dehydrated conditions was first irradiated with γ -rays at a dose of 10 kGy and then
 187 washed with a solution of 5 wt.% LiCl/DMAc to remove any residues of unreacted
 188 cellulose, degradation products, and EMI-acetate. However, the gel products were not
 189 retained in the process. It has been reported that the radiation-induced reduction of
 190 chlorophenols in RTILs can occur effectively under dehydrated conditions at a dose of
 191 20 kGy (Kimura et al., 2008, 2010). In the present case, the amount of
 192 radiation-induced species was not considered sufficient to cause the cross-linking
 193 reaction of cellulose. Therefore, water, which is a source of reactive species such as

194 hydroxyl radicals and hydrated electrons produced by ionizing radiation, was added to
195 the dried cellulose solution to enhance the radiation-induced cross-linking of cellulose.
196 The cellulose EMI-acetate solution with a water content of 18 wt.%, was prepared
197 under humidity-controlled air conditions and left overnight to form a physical gel. The
198 gel thus obtained could be completely dissolved in a 5 wt.% LiCl/DMAc solution. The
199 physical gel was irradiated with γ -rays at a dose of 10 kGy, followed by washing with a
200 5 wt.% LiCl/DMAc solution to obtain an insoluble residue.

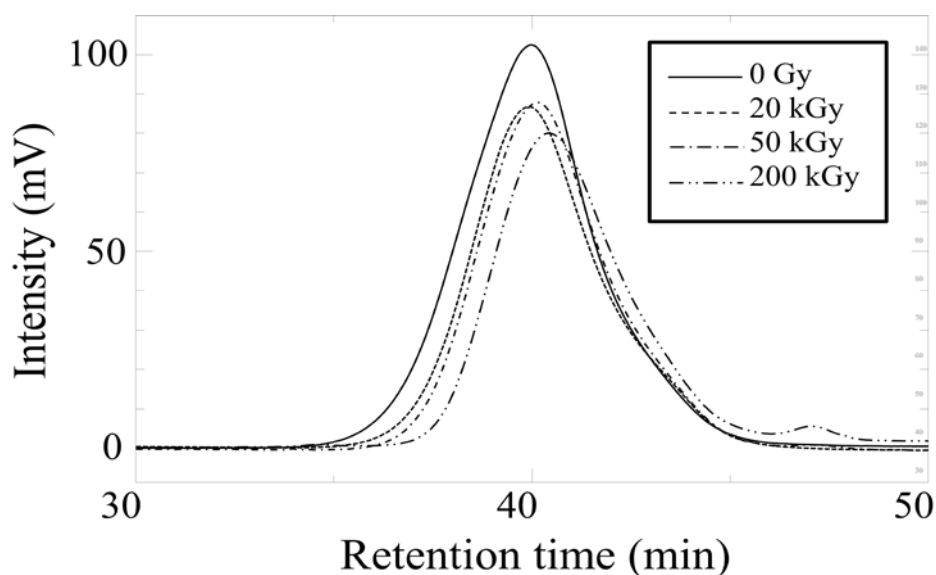
201 The physical properties of the insoluble product thus obtained were evaluated by
202 dynamic viscoelasticity measurements (Techwipharat et al. 2008; Zhou et al. 2008).
203 The interactions among the cellulose chains of the physical gel are weak and reversible,
204 and the gel reverts to a sol at temperatures above 353 K. Figure 1 shows the storage
205 modulus (G') and loss modulus (G'') of the solution containing 20 wt.% of cellulose in
206 EMI-acetate before and after irradiation measured at 353 K. The G' and G'' values of
207 the as-prepared physical gel (before irradiation) increase with frequency, without
208 showing the crossover point of G' and G'' . This indicates that the as-prepared sample
209 (before irradiation) is a sol at 353 K (Song et al., 2011). On the other hand, after
210 irradiation, the G' curve of the insoluble product obtained by irradiation at a dose of 20
211 kGy was almost constant, while that of the G'' increased with frequency. This implies

212 that the irradiated sample is a gel at 353 K. Based on these results, the obtained
213 insoluble product can be considered as a cellulose gel.



214
215 **Figure 1** Dynamic viscoelasticity measurement of 20 wt.% cellulose in EMI-acetate,
216 containing 18 wt.% of water at 353 K under aerated conditions, before and after γ -ray
217 irradiation at 20 kGy.

218
219 The cellulose gel fraction was separated from the γ -ray irradiated solution of 20
220 wt.% cellulose dissolved in EMI-acetate with 18 wt.% of water at 298 K under aerated
221 conditions. The soluble part, i.e., the sol, was evaluated using the GPC system with the
222 refractive index detector. The peak retention time of 40.0 min could be assigned to the
223 cellulose before irradiation. However, with increase in irradiation dose, the retention
224 times slowly shift towards higher values, as shown in Figure 2.



225

226 **Figure 2** Gel permeation chromatograph of the solution of 20 wt.% cellulose
 227 dissolved in EMI-acetate with 18 wt.% of water at 298 K under aerated conditions,
 228 compared with a neat cellulose gel at doses of 0 Gy (line), 20 kGy (dot line), 50 kGy
 229 (chain line), and 200 kGy (two-dot chain line).

230 For the absorbed dose of up to 20 kGy, the intensity of the peak at 40.0 min was clearly

231 reduced by approximately 13 %. This could be attributed to the fact that the irradiation

232 products mainly consist of the cellulose gel and degradation products. The

233 weight-average molecular weight of the cellulose samples at the dose of 0, 20, 50, and

234 200 kGy was found to be 1.3×10^5 , 1.1×10^5 , 1.0×10^5 , and 8.8×10^4 g·mol⁻¹,

235 respectively, as evaluated using the PMMA standard solution. As mentioned earlier, it

236 has been reported that the degree of polymerization and the weight-average molecular

237 weight of cellulose dissolved in BMI-chloride are reduced by approximately 50 % at

238 the dose of 200 kGy (Hao et al. 2012). In the present study, the rate of decrease in the

239 weight-average molecular weight of cellulose dissolved in EMI-acetate was found to

240 be 32 % at the same dose of 200 kGy. This is lower than that reported for cellulose
241 dissolved in BMI-chloride. The intensity of the peak at the retention time of 47.0 min
242 increased when the dose was greater than 20 kGy (Figure 2). This peak can be assigned
243 to the irradiation products corresponding to the weight-average molecular weight of
244 $7.1 \times 10^3 \text{ g}\cdot\text{mol}^{-1}$.

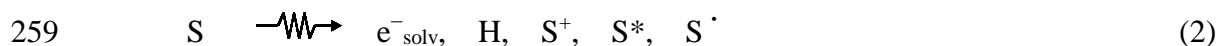
245

246 *3.2 Mechanism underlying the formation of cellulose gels in RTILs*

247 In general, the solid cellulose is decomposed by ionizing radiation (Charlesby et
248 al. 1957; McLaren 1978; Kumakura et al. 1979; Charlesby 1995; Takacs et al. 1999;
249 Driscoll et al. 2009). Upon γ -ray irradiation, radicals emerge preferentially as a result
250 of the fracture of C-H bonds in the weakened 1 and 4 positions of cellulose.
251 Subsequently, the direct cleavage of the chemical bonds occurs at these positions in the
252 polymer chains to produce cellulose of lower molecular weight (Yershov et al. 1977;
253 Hao et al. 2012). However, in this study, cellulose gel was produced when the solution
254 of 20 wt.% cellulose dissolved in EMI-acetate was irradiated under humid conditions
255 without cross-linking agents.

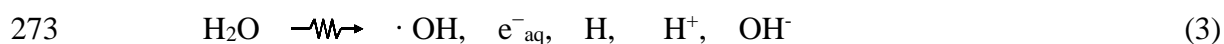
256 Reactive intermediates, such as solvated electrons, hydrogen atoms, positive ions,
257 excited molecules, and radicals, are formed in the organic solvent (Woods et al.,

258 1994.).



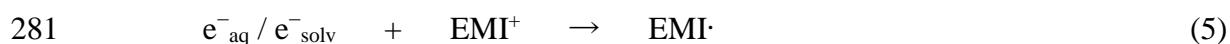
260 where S, e^-_{solv} , and H are represent solvent, solvated electrons, and hydrogen atoms,
261 respectively. The rate constants corresponding to the reaction of radiation-induced
262 reductive species, such as solvated electrons, with some aromatic and halogenated
263 organic solutes is typically higher than the corresponding diffusion-controlled rate
264 constants in RTILs under dehydrated conditions (Wishart et al., 2003; Wishart et al.,
265 2005; Kimura et al., 2008; Kimura et al., 2010). The cellulose gel, however, was not
266 produced in EMI-acetate under dehydrated conditions. The solvated electrons have low
267 reactivity toward cellulose as a saturated hydrocarbon, and are not related to the
268 radiation-induced cross-linking of cellulose under the conditions.

269 The cellulose EMI-acetate solution having 18 wt.% of water was irradiated with
270 γ -rays to produce the cellulose gel as stated above. In this reaction, water is
271 decomposed by the ionizing radiation, resulting in the formation of a hydroxyl radical,
272 hydrated electron, hydrogen atom, proton and hydroxide ion (Getoff et al., 1996).

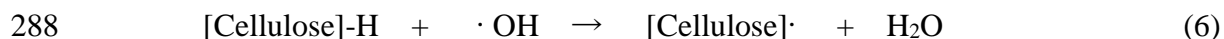


274 The hydroxyl radicals, in particular, contribute to the production of cellulose gel,
275 owing to their high reactivity toward saturated hydrocarbons and high yield upon water

276 radiolysis (Buxton et al., 1988). Besides, the hydroxyl radicals, hydrated electrons
277 (e^-_{aq}) and the electrons solvated by the EMI-acetate (e^-_{solv}) also react with the
278 alkyimidazolium of the EMI-acetate to form alkyimidazolium radicals (Behar et al.,
279 2001; Wishart et al., 2005).

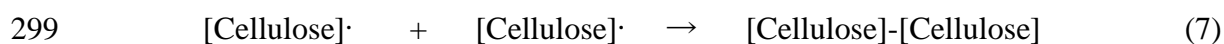


282 As mentioned earlier, γ -ray irradiation of the solution of 20 wt.% cellulose dissolved in
283 EMI-acetate under dehydrated condition did not result in the formation of gel.
284 Therefore, the solvated electrons and alkyimidazolium radicals cannot be related to
285 the radiation-induced cross-linking of cellulose. On the other hand, hydroxyl radicals
286 are considered to oxidize the carbon atoms of cellulose to produce cellulose radicals
287 (Isogai et al., 1998).



289 There is a possibility that the cellulose radicals are formed in the solid phase or in
290 another organic solvent upon γ -ray irradiation. However, the macro-sized cellulose
291 radicals would not diffuse easily in the solid phase. Besides, the solubility of cellulose
292 is less than 10 wt.% in organic solvents, such as chloroform in dimethylsulfoxide,
293 *N*-methylmorpholine-*N*-oxide, LiCl in DMAc, and calcium chloride in methanol. In

294 contrast, the radiation-induced cellulose radicals in EMI-acetate readily diffuse more
295 easily than that in the solid phase to facilitate cross-linking reaction, and solubility of
296 cellulose in EMI-acetate is higher than that in the organic solvent systems listed. Based
297 on these observations, following reaction paths are proposed for the formation of the
298 cellulose gel.



300 The cellulose having radicals at the 6th carbon atom is expected to participate in the
301 cross-linking reaction because the side chain of the polysaccharide derivatives is
302 considered to be the cross-linking position (Saiki et al., 2010; Saiki et al., 2011).

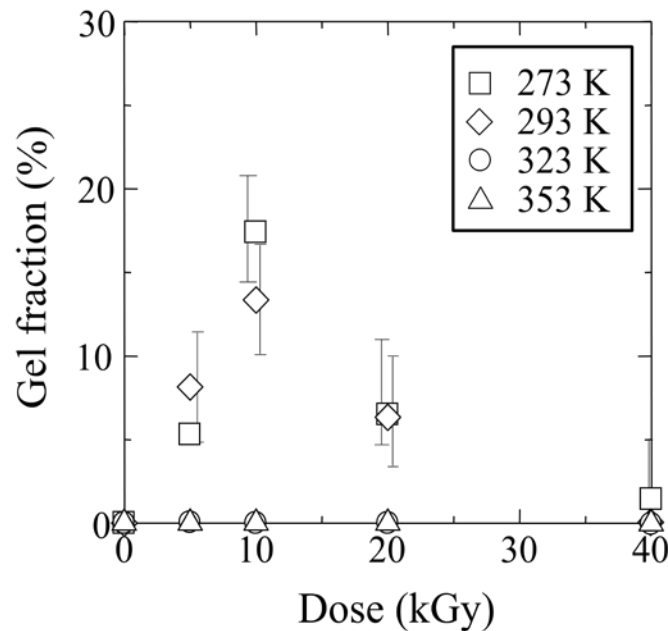
303

304 *3.3 Gel fraction of cellulose in RTILs under several conditions*

305 It has been reported that structure of the cellulose polymer chains in EMI-acetate
306 tends to change as a function of the concentration of cellulose and the temperature of
307 γ -ray irradiation (Song et al., 2011). This effect was further analyzed in this study, as
308 the yield of the obtained cellulose gel was found to depend on the oriented structures
309 of the cellulose polymer chains. The yield of the gel obtained by γ -ray irradiation of
310 cellulose dissolved in EMI-acetate was estimated in terms of the gel fraction by the
311 following equation:

312
$$\text{Gel fraction (\%)} = (W/W_0) \times 100 \quad (8)$$

313 where W_0 and W are the initial mass of the dried sample before irradiation and the final
314 mass of the insoluble product obtained after irradiation, washing, and drying,
315 respectively. The gel fraction of the cellulose gel in EMI-acetate having 18 wt.% of
316 water under air-saturated conditions increased with increase of dose at 298 K, and
317 reached a maximum value of 17 % at the dose of 10 kGy, as shown in Figure 3.

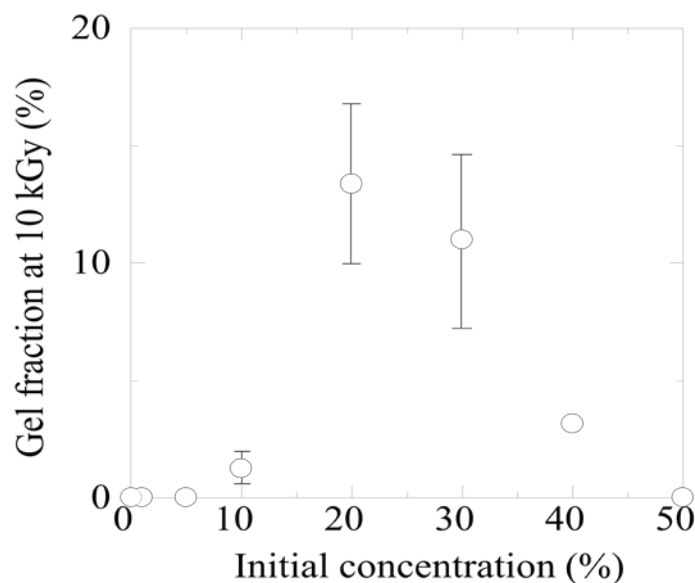


318
319 **Figure 3** Gel fraction of 20 wt.% cellulose in EMI-acetate containing 18 wt.% of water
320 under aerated conditions plotted as a function of the absorbed dose.

321 The gel fraction does not exist at temperatures above 323 K. The physical gel begins to
322 revert to a sol above 323 K (Song et al., 2011), and becomes a solution at 353 K under
323 dehydrated condition. For the solution with lower viscosity, the scission reaction is
324 dominant by ionizing radiation (Nagasawa et al. 2000). Therefore, the optimal

325 production of the cellulose gel was obtained under the physical gel condition at
326 temperatures less than 298 K. In addition, a paste-like solution of 20 wt.% cellulose in
327 DEMA-formate with 18 wt.% of water under air-saturated conditions at 298 K was
328 also irradiated at a γ -ray dose of 10 kGy. This resulted in the formation of cellulose gel
329 with a yield of 19 %. However, the formation of cellulose gel was not obtained in other
330 RTILs in the presence of water, as listed in Table 1. This could be attributed to the fact
331 that the cellulose was heterogeneously dispersed in these RTILs, as opposed to that in
332 the EMI-acetate and DEMA-formate.

333 Furthermore, the cellulose at an initial concentration of 1–50 wt.% in EMI-acetate
334 having 18 wt.% of water under air-saturated conditions at 298 K was also irradiated at
335 a dose of 10 kGy. For initial concentration higher than 40 wt.%, the cellulose was
336 found to disperse heterogeneously in EMI-acetate. The gel fraction of the obtained
337 cellulose gel is illustrated in Figure 4. The formation of cellulose gel is effective in the
338 initial concentration range of 20–30 wt.%. On the other hand, the cellulose solutions
339 had low gel fractions for their initial concentrations either lower than 10 wt.% or
340 higher than 40 wt.%.

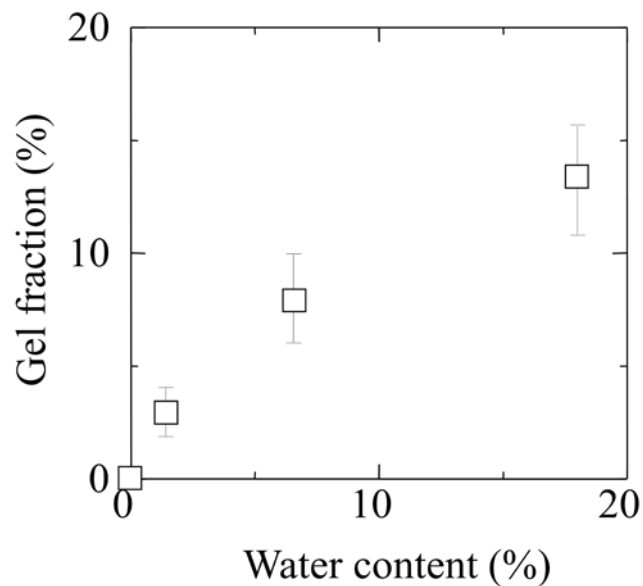


341 **Figure 4** Initial concentration dependence of the gel fraction of cellulose in
 342 EMI-acetate having 18 wt.% of water at 298 K under aerated conditions at 10 kGy.
 343
 344 In general, when polymers are irradiated with ionizing radiation, both the cross-linking
 345 and scission reactions occur simultaneously. The scission reaction is dominant over the
 346 cross-linking reaction when the concentration of polysaccharides in the organic solvent
 347 is less than approximately 10 wt.% (Tabata et al., 1991; Nagasawa et al., 2004).
 348 However, irradiation of the samples in the paste-like state is effective to facilitate the
 349 cross-linking of other polysaccharide derivatives (Fei et al., 2000; Zhao et al., 2003;
 350 Nagasawa et al., 2004). Besides, a paste-like solution of cellulose dissolved in
 351 EMI-acetate can be prepared for the concentration range of 20–30 wt.%. In the present
 352 study, the ratio of degradation to cross-linking density (p_0/q_0) was estimated using the
 353 Charlesby–Rosiak equation (Olejniczak et al., 1991):

354
$$s + s^{1/2} = \frac{p_0}{q_0} + \left(2 - \frac{p_0}{q_0}\right) \frac{D_v - D_g}{D_v - D} \quad (9)$$

355 Here, s , D , D_g , and D_v are the sol fraction, absorbed dose, gelation dose, and virtual
356 dose, respectively. For the solution of 20 wt.% of cellulose in EMI-acetate at 298 K
357 with 18 wt.% of water under air-saturated conditions, the value of p_0/q_0 was estimated
358 to be 1.86. The value implies that the scission reaction occurs more effectively than the
359 cross-linking reaction, resulting in a low cellulose gel yield of 17 %. The p_0/q_0 values
360 for 10, 30, and 40 wt.% of cellulose in EMI-acetate at 298 K having 18 wt.% of water
361 under air-saturated conditions were estimated to be 1.97, 1.74, and 1.92, respectively.

362 The respective effects of the water content on the gel fraction of cellulose in
363 EMI-acetate were also investigated by γ -ray irradiation. The gel fraction of cellulose
364 increases with water content, as shown in Figure 5. As mentioned earlier, the
365 radiation-induced cross-linking of cellulose is considered to be initiated by the
366 hydroxyl radicals. The sample solution containing 34 wt.% of water, which was
367 partially clouded, was irradiated with γ -rays. However, cellulose gel was not obtained
368 under this condition. The crystallization of the cellulose became more pronounced with
369 decrease in the concentration of the RTILs in the solution. The cellulose solution
370 containing 34 wt.% of water is considered to be the similar to that of the solid cellulose.
371 Hence, the scission reaction of cellulose would have been dominant.



372
 373 **Figure 5** Water content dependence of the gel fraction obtained by γ -ray irradiation of
 374 20 wt.% cellulose in EMI-acetate at 298 K under aerated conditions at 10 kGy.
 375

376 *3.4 Electronic conductivity of cellulose gel*

377 Recently, polymer ion gels that are swollen with RTILs have been applied as
 378 electrolytes in dye-sensitized solar cells (Ueno et al., 2008), electroactive polymer
 379 actuators (Imaizumi et al. 2012), and thin-film transistors (Lee et al., 2007), owing to
 380 their excellent chemical and thermal stabilities, wide potential windows, and high
 381 electrical conductivities of 10^{-2} – 10 $\text{mS}\cdot\text{cm}^{-1}$, which depend on the type of RTIL used
 382 in the process (Kitazume et al., 2005; Lodge, 2008). In this study, cellulose gels were
 383 swollen with water, methanol, ethanol, dichloromethane, DMAc, EMI-acetate, and
 384 DEMA-formate. The swelling ratio of cellulose gel in EMI-acetate was 3.7 g swollen
 385 gel/g dried gel. The electronic conductivity of the cellulose gel containing

386 EMI-acetate, as estimated by the chemical impedance method at 298 K was found to
387 be $3.0 \text{ mS}\cdot\text{cm}^{-1}$. This is sufficiently higher than the required electronic conductivity
388 of $0.1 \text{ mS}\cdot\text{cm}^{-1}$ for the organic solid electrolytes (Ohno, 2006). The conductivity of
389 EMI-acetate was the same with that of the cellulose gel containing them. Therefore,
390 cellulose gel has great potential for applications in optoelectronic and medical devices
391 as a fascinating biopolymer and sustainable raw material.

392

393 **4. Conclusion**

394 For the first time in this study, we have demonstrated the possibility of
395 producing cellulose gels by γ -ray irradiation of cellulose in RTILs under humid
396 conditions. Water was found to be essential for the formation of the cellulose gel, as
397 the hydroxyl radicals formed by the radiolysis of water play an active role in the
398 cross-linking of cellulose. RTILs having hydrophobic imidazolium/ammonium and
399 proton-accepting carboxylate groups were found to be suitable mediums for the
400 production of cellulose gels. The gel fraction could be controlled by adept regulation of
401 following conditions: initial concentration of cellulose, irradiation temperature, water
402 content, and the absorbed dose. The cellulose gel thus obtained had an electronic
403 conductivity of $3.0 \text{ mS}\cdot\text{cm}^{-1}$, showing great potential for applications in optoelectronic

404 and medical devices.

405

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